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Patrice Tankam,^{a,b} Anand P. Santhanam,^c Kye-Sung Lee,^{a,d} Jungeun Won,^e Cristina Canavesi,^f and Jannick P. Rolland^{a,b,e,f,*}

^aUniversity of Rochester, The Institute of Optics, 275 Hutchinson Road, Rochester, New York 14627

^bUniversity of Rochester, Center for Visual Science, 601 Elmwood Avenue, Rochester, New York 14642

^cUniversity of California, Department of Radiation Oncology, Los Angeles, 200 Medical plaza drive, Los Angeles, California 90095

^dKorea Basic Science Institute, Center for Analytical Instrumentation Development, Daejeon 305-806, South Korea

^eUniversity of Rochester, Department of Biomedical Engineering, 275 Hutchinson Road, Rochester, New York 14627

¹LighTopTech Corp., 150 Lucius Gordon Dr., Ste 115, West Henrietta, New York 14586

Abstract. Gabor-domain optical coherence microscopy (GD-OCM) is a volumetric high-resolution technique capable of acquiring three-dimensional (3-D) skin images with histological resolution. Real-time image processing is needed to enable GD-OCM imaging in a clinical setting. We present a parallelized and scalable multi-graphics processing unit (GPU) computing framework for real-time GD-OCM image processing. A parallelized control mechanism was developed to individually assign computation tasks to each of the GPUs. For each GPU, the optimal number of amplitude-scans (A-scans) to be processed in parallel was selected to maximize GPU memory usage and core throughput. We investigated five computing architectures for computational speed-up in processing 1000×1000 A-scans. The proposed parallelized multi-GPU computing framework enables processing at a computational speed faster than the GD-OCM image acquisition, thereby facilitating high-speed GD-OCM imaging in a clinical setting. Using two parallelized GPUs, the image processing of a $1 \times 1 \times 0.6$ mm³ skin sample was performed in about 13 s, and the performance was benchmarked at 6.5 s with four GPUs. This work thus demonstrates that 3-D GD-OCM data may be displayed in real-time to the examiner using parallelized GPU processing. © *The Authors. Published by SPIE under a Creative Commons Attribution 3.0 Unported License. Distribution or reproduction of this work in whole or in part requires full attribution of the original publication, including its DOI. [DOI: 10.1117/1.JBO.19.7.071410]*

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1 Introduction

Noninvasive skin imaging is envisioned to play a prevalent role in the future of dermatology. Because of its accessibility, skin is amenable to noninvasive diagnostics and therapeutic monitoring; an additional important advantage offered by noninvasive skin imaging is timeliness, resulting in a more effective use of the clinician's time spent treating a patient. Extensive research has been conducted in the past few decades toward developing noninvasive skin imaging and diagnostic modalities. Optical coherence tomography (OCT) offers the capability to image layers and structures well below the surface of the skin.¹⁻ Numerous studies using OCT have been performed to image subsurface layers and structures of skin, including the epidermis, dermal-epidermal junction, dermis, hair follicles, blood vessels, and sweat ducts.⁶⁻⁹ Clinical studies suggest that OCT may be useful for noninvasive diagnosis of skin diseases and to assess wound healing.^{2-5,10-17} The lateral resolution of conventional OCT instruments is limited to tens of micrometers, hampering the adoption of OCT in a wide range of applications that require cellular resolution comparable to or approaching histological resolution. The numerical aperture (NA) of the optics sets the lateral resolution in the focal plane of the optics and throughout the depth of focus. The depth of focus is inversely proportional to the NA. As a result, OCT typically operates at low NA of around 0.05 to 0.025 with a corresponding lateral resolution in the order of 10 to 20 μ m, which enables a large depth of focus on the millimeter scale (0.6 to 2.4 mm). Various hardware and software methods have been investigated to address the trade-off between lateral resolution and depth of focus, including axicon lenses to generate Bessel beams or phase masks, holoscopy, and computational techniques. Bessel beams have successfully demonstrated imaging in biological tissue with lateral resolution ranging from ~ 5 to 8 μ m, with extended depth of focus in the millimeter range.¹⁸⁻²⁰ The main limitation of Bessel beams imaging has been the reduced light efficiency of these systems. Phase masks have been investigated to create an annular mask²¹ or add spherical aberration to the optical system,²² with a factor of \sim 2 to 3 improvement in the depth of focus at the expense of some loss of image quality throughout the range. Holoscopy, which combines full field Fourier-domain OCT and numerical reconstruction of digital holography, has been introduced as a solution to achieve extended depth of imaging with constant sensitivity and lateral resolution. The lateral resolution is not limited by the NA, but rather by the numerical reconstruction distance of the holograms. Depth of imaging of \sim 3 mm and lateral resolution of ~10 μ m was reported.²³ However, holoscopy suffers from noncompensated phase error caused by multiple scattering (non-

^{*}Address all correspondence to: Jannick P. Rolland, E-mail: rolland@optics .rochester.edu.

ballistic) photons in highly scattering samples.²³ Computational methods such as three-dimensional (3-D) Fourier-domain resampling have been demonstrated in combination with interferometric synthetic aperture microscopy to extend the imaging depth to 1.2 mm in skin *in vivo*; these techniques require an accurate and stable phase measurement.²⁴

Optical coherence microscopy (OCM) was introduced to achieve cellular resolution using a higher NA objective (i.e., ~ 0.2) than conventional OCT (i.e., ~ 0.04); the gain in resolution in OCM is reached at the expense of a limited depth of focus in the order of 100 to 200 μ m.²⁵ Gabor-domain optical coherence microscopy (GD-OCM) was proposed by our group in 2008 to dynamically extend the imaging depth of OCM²⁶ and has since started to be adopted in other research groups as well.²⁷ A liquid lens is dynamically refocused at different depth locations to acquire multiple images that are then combined in a single volume. The custom optics can refocus up to a 2-mm imaging depth. In skin, an imaging depth of about 0.6 mm was achieved with an invariant lateral resolution of 2 μ m throughout the volume.²⁸ Provided the increased lateral resolution by an order of magnitude compared with conventional OCT, the images acquired with GD-OCM have a shallower depth of focus around the focal plane controlled by the liquid lens, typically in the order of 60 to 100 μ m.²⁹ Therefore, in order to image a volume up to 0.6 mm in depth for example, the liquid lens is dynamically refocused six times to image six volumes, which are then fused in postprocessing to produce a volumetric image with $2-\mu m$ resolution throughout the 0.6-mm depth.³⁰ The large dataset and the multiple computational tasks associated with GD-OCM compound the need for fast processing; performing the processing steps on conventional architectures takes about two orders of magnitude longer than the acquisition steps. Critical to the adoption of GD-OCM in the clinical workflow is a fast, real-time processing, and rendering of the high-resolution images.

Recently, graphics processing units (GPUs) were shown to be powerful tools for general numerical simulation,^{31–35} signal processing,³⁶ and image processing for a variety applications.³⁷ The use of GPUs has been investigated for several imagingrelated tasks, ranging from image processing steps to imagebased modeling and clinical intervention guidance.³⁸ The GPU technology was also reported to solve computational problems related to medical imaging.^{39,40} Several studies conducted in the last 4 years have shown how GPUs can improve the processing speed of OCT imaging.41-55 The use of multiple GPUs for OCT has also been investigated by peers. Huang et al.⁵⁶ demonstrated the use of dual GPUs to simultaneously compute the structural image intensity and phase Doppler imaging of blood flow on both a phantom and the chorioallantoic membrane. The authors reported a frame rate of 70 fps with an image size of 1000×1024 pixels. The same authors demonstrated in a different paper GPU-based motion compensation of handheld manual scanning OCT.57 Zhang and Kang also investigated the use of dual GPUs architecture to speed up the processing and rendering steps of an OCT system designed to guide micromanipulation using a phantom model and vitreoretinal surgical forceps. The first GPU was dedicated to data processing, whereas the second was used for rendering and display. A volume rate of 5 volumes per second with the volume size of $250 \times 98 \times 1024$ voxels was reported.⁵⁸ Later, the same group demonstrated the use of dual-GPU architecture to guide microsurgical procedures of microvascular anastomosis of the rat femoral artery and ultramicrovascular isolation of the retinal arterioles of the bovine retina.⁵⁹ A display rate of 10 volumes per second for an image size of $160 \times 80 \times 1024$ voxels was reported. In these recent advancements with multiple GPUs, two GPUs were considered, where one GPU was typically dedicated to processing and another GPU was used for image rendering.

Compared with OCT, GD-OCM faces additional challenges deriving from the higher imaging resolution of 2 μ m that imposes 1- μ m sampling, which results in a significantly larger dataset to be processed. Also, the computation and fusing of six volumes of data is a demanding task. We propose here a parallel computational framework using multiple GPUs to enable real-time imaging capabilities of GD-OCM. In the following sections, we review the architecture of the GD-OCM system and detail the imaging process in the central processing unit (CPU) in order to identify the computational bottleneck of the imaging process. We then describe the proposed parallelized GPU framework to overcome the limitations (See Sec. 3).

2 Methods

2.1 System Description

The current GD-OCM system fits on a movable cart. The handheld scanning probe is attached to an articulated arm that can be easily adjusted to fit the region of the skin that the clinician wants to image. The imaging system has micron-class resolution of 2 μ m in skin tissue (average refractive index of 1.4), both axially and laterally. The light source is a superluminescent diode laser centered at 840 nm with 100 nm FWHM (BroadLighter D-840-HP-I, Superlum®, Ireland). The microscope objective probe with 2-mm field-of-view incorporates a liquid lens, which allows dynamic-focusing in order to image different depths of the sample while maintaining a lateral resolution of 2 μ m within the imaging depth of up to ~2 mm by design.^{28,29} A custom dispersion compensator and a custom spectrometer with a high-speed CMOS line camera (spl4096-70 km, Basler Inc., Exton, Pennsylvania) are used to acquire the spectral information.^{60,61} With a set depth of focus of 100 μ m, the liquid lens is refocused six times to image 600 μ m in depth in skin tissue, yielding six volumes of data to acquire and to process. Conventionally, a skin sample of $1 \times$ $1 \times 0.6 \text{ mm}^3$ acquired with GD-OCM generates 49 GB (i.e., 6 zones \times 1000 \times 1000 \times 4096 \times 2 bytes) of data to be processed. Although the current total acquisition time is 1.43 min, the image processing steps on this high-resolution data may take up to 3.5 h on a conventional sequential architecture, as discussed in the following section, before the scanned data can be visualized in 3-D by a clinician.

2.2 Description of the Imaging Process in CPU

The imaging platform runs on a workstation equipped with a Supermicro X8DTG-QF motherboard. The system operates on 64 bit Microsoft[®] Windows[®] 7 with two processors Intel[®] core i7/Xeon (X5650 6-core 2.66-GHz CPUs, each with 12-MB cache) and 48 GB of RAM to allow the operating system to run smoothly while the GD-OCM images are being acquired. High-efficiency data security and storage on a 2.4-TB (eight Seagate 300GB SAS 15K hard drives) hard drive is made possible by an LSI, San Jose, California MegaRAID 9260 redundant array of independent disks (RAID) card. A Camera-link Bitflow frame grabber Karbon-CL, Woburn, Massachusetts is used to buffer the data from a camera, and a DAQ card (National Instruments, PCI-6733) is dedicated to control the galvanometer scanners and liquid lens and generate the trigger signal for the camera to synchronize the scanning and the acquisition at the beginning of each zone. In CPU-based processing, the imaging process consists of acquisition, postprocessing, fusing, and rendering steps. All these steps, excluding rendering, run on LabVIEWTM 2012 software (National Instruments, Austin, Texas).

2.2.1 Acquisition

The acquisition step consists of lateral scanning of the same skin area six times with different focal lengths of the liquid lens. For each focal length, 1000×1000 amplitude scans (A-scans) spectra with a lateral sampling interval of 1 μ m are acquired. Data with single precision are saved in parallel on the hard drive in binary format. In order to run acquisition and saving independently, a buffer was created in LabVIEWTM to hold the acquired data while saving is in progress using the high-speed storage capability of RAID technology. After the acquisition of the first zone, the focal length is shifted to the next zone and the lateral scanning is repeated. After acquisition of all six zones, six 1000×1000 A-scans are saved on the disk. The acquisition uses a high-speed CMOS line camera with 70,000 A-scans/s, for an acquisition time of 14.3 μ s per A-scan. The total acquisition time is then 1.43 min for the six zones. A further reduction in acquisition is anticipated with the higher frame rate cameras that have already reached the market. Each A-scan consists of a binary spectrum of 4096 pixels, thus the acquisition step generates 49 GB (6 zones \times 1000 \times 1000 \times 4096 \times 2 bytes) of data that is saved to the disk for postprocessing.

2.2.2 Postprocessing

After reading data from the disk, the postprocessing steps consist of performing DC removal, k-space linearization, fast-Fourier transform (FFT), gray scaling, and auto-synchronization.

The DC term for each B-scan is a one-dimensional (1-D) array of 4096 pixels, which is obtained by computing the mean of the 1000 A-scans that form the B-scan. The DC term is then removed from each A-scan. Next, each A-scan undergoes k-space linearization, FFT, and gray level and log scaling using a modified version of existing functions in LabVIEWTM. To achieve high-speed acquisition, the hardware synchronization is applied just once at the beginning of each zone. As a consequence, a drift in hardware synchronization between the camera and the scanning is experienced. An auto-synchronization

software was developed and implemented to compensate the drift with an intercorrelation algorithm applied between two consecutive B-scans. The peak of the intercorrelation corresponds to the number of shifted A-scans. The region of interest of the shift-compensated frames is buffered in the memory for the fusing step. Each zone has a dedicated buffer to facilitate the fusing step. The total postprocessing time, including reading data, is 29 min for one zone, yielding 2.9 h for six zones (see detailed timing in Table 1).

2.2.3 Fusing

In the fusing step, the six zones of the sample are fused in one volume using the Gabor fusion technique.^{27,62} For each B-scan, the six frames are accessed from the buffers; only the focused region (the region within $\pm 50 \ \mu$ m around the focal plane) of the six frames contributes to the final image. Thus, each frame is multiplied by a window of width ~100 μ m centered at the focal plane of the dynamic focus probe. The window serves as a weighting function for each frame in the fusing process. The six windows are preoptimized based on the voltage applied to the liquid lens and the focal shift for each acquired image. The final image is obtained by adding the six windowed frames. All focused B-scan frames are saved back to the disk using binary format. The computation time for the fusing process is estimated at 20 min, accounting for the disk saving time.

2.2.4 Rendering

Voxx and open source ImageJ are used to render the volumetric image and display the two-dimensional (2-D) and 3-D images. The time needed for rendering is ~ 10 min.

In summary, the 3-D imaging and visualization of a $1 \times 1 \times 0.6 \text{ mm}^3$ sample using a sequential implementation takes about 3.5 h. The analysis of the sequential implementation (Table 1) shows that the k-space linearization is the most time-consuming operation (44%); followed by the gray level and log scaling (29%); fusing (8%); DC removal (8%); FFT (7%). Saving, loading, and auto-synchronization account for 4% altogether.

As a next step to increase the computational speed on CPU, the use of pipelined computation was investigated, in which the operations with independent parameters are regrouped in different operation blocks, leveraging more advanced multithread CPU capabilities. Specifically, a process pipelining approach was employed, in which different operation blocks in the postprocessing and fusing steps were separated and performed in a pipelined manner. Figure 1 shows the flowchart of the proposed pipelined computation architecture in which the two

Table 1 Timing of the processing steps and computational speed-up between sequential and pipelined implementations.

1 Zone (4096 × 1000 × 1000	Loading data from the disk	DC removal	K-space linearization	Fast Fourier transform (FFT)	Gray level and log scaling	Auto synchronization	Fusing	Saving	Total time (s)
Sequential central processing unit (CPU) time(s)	30	148	846	137	553	5	150	54	1923 (32 min)
Pipelined CPU time (s)	Running in parallel with the next operation block	148	846	137	Running in para	allel with previous	operatio	n block	1131 (19 min)



Fig. 1 Pipelined computation in central processing unit (CPU).

most time-consuming operations are separated in two different blocks running in parallel.

In the first block, the input A-scan data are loaded into a queue data structure while they are also simultaneously accessed for image processing in the second operation block, which consists of three sequential steps—DC removal, k-space linearization, and FFT. Similarly, another queue data structure is used to hold the modulus of the FFT outputs. The last operation block, dedicated to log scaling, auto-synchronization, and fusing, accesses the queue in a parallel manner and saves the final fused data into the hard drive. Table 1 summarizes the computational speed-up of the pipelined approach as compared with the sequential implementation.

The pipelined CPU implementation completes processing one zone in ~19 min, offering a $1.7 \times$ speed-up over the 32 min of the sequential implementation. In this computation, as in prior work, k-space linearization is the bottleneck. In prior investigations, hardware solutions to k-linearization have been reported with good results.^{63–65} However, regardless of the approach to addressing the bottleneck, real-time imaging on CPU cannot be achieved, even with the pipelined approach. The CPU implementation is fundamentally limited by the number of cores compared with GPU that can allow more advanced parallelization implementation for the entire processing.

2.3 Proposed Multi-GPU Framework for GD-OCM

Since the 1-D A-scan signals can be processed independently of each other, a parallelized scalable processing can fully leverage a multi-GPU system. The multi-GPU framework was designed to achieve the acquisition of each zone in parallel with the processing of the previous zone and yields the 3-D visualization of the sample within seconds after the acquisition of the entire volume is completed.

The motherboard of the workstation (Supermicro[®]) can hold up to four GPUs and was configured based on the requirement to complete the processing of one zone within seconds, while the next zone is being acquired. Each of the GPUs is connected to the main system using a PCIe-2.0 connectivity facilitating up to 8-GB/s data transfers between the CPU and each of the GPUs. The GPU processes run independently of each other and can occasionally communicate with each other using the PCIe-2.0 connectivity, with the CPU forming the intermediary communication step. Buffers are used, as in the CPU implementation, to serve as temporary memory. After the processing of each frame, only the focused region $(1000 \times 100 \ \mu m^2)$ is held in the GPU buffer, thus dividing by six the size of data to be managed during the fusing step.

Figure 2 shows a schematic of the GD-OCM instrument and the proposed architecture of the multi-GPU-based GD-OCM imaging system. The acquisition time for each of the six zones, accounting for the focal length of the liquid lens being adjusted six times, is 14.3 s. During the acquisition of the first zone, which consists of a data size of 1000×1000 A-scans, the data is held in a temporary CPU buffer. When the acquisition of this zone is completed, the data is structured and divided into four sections, which are transferred to the four GPUs via the four PCIe-2.0 interfaces. During that time, the focal length of the liquid lens is shifted by 100 μ m to acquire the next zone. The acquisition of the next zone is done in parallel with the processing of the previous zone. The processing consists of DC removal, k-space linearization, computer unified device architecture (CUDA) FFT, gray level and log scaling, and auto-synchronization. Once the processing is completed for each frame, the resultant data is windowed to retain only the focused region. Data is held in temporary buffers in each GPU to be fused with the next zone. The GPUs are then released to handle processing of the next available zone. After the processing of each zone, the focused region is fused with the previous fusing result and hold in GPU memory as illustrated in Fig. 3 for one B-scan (1000 A-scans). Once all six zones are processed, visualization of the 3-D scan is enabled by GPU-based volume rendering, as well as 2-D visualization. Visualizing the 3-D structure in real-time is a computationally complex task mainly because of the size of the 3-D data $(1000 \times 1000 \times 400 \text{ voxels})$. For this study, the rendering was performed with GPU-based real-time volume rendering in one of the GPUs, providing a rendering time of 50 ms for one volume.

2.4 Implementation of the Multi-GPU Framework

The software development framework for the proposed system consists of LabVIEW™ 2012 and NVIDIA® CUDA 5.0 programming interfaces. LabVIEWTM was employed to interface with the GD-OCM image acquisition system and to control access to the GPU cards. Communication between LabVIEWTM and CUDA was achieved with a dynamic-link library (DLL) developed in Microsoft® Visual Studio 2010. The different steps of the processing, including DC removal, k-space linearization, gray scaling, and auto-synchronization, were implemented in C++ using Microsoft® Visual Studio 2010; CUDA Fast Fourier Transform (CUFFT) was performed with the existing function in the CUDA environment. Parallelized access to the GPUs was made possible via multiple DLL calls with parameters such as the GPU identity, calibration, and frame-related parameters. Although each DLL call was designed to employ multiple cores in parallel, a set of DLL calls was implemented in parallel to fully employ the GPU cores available in each GPU and initiate the GPU boost capability.

3 Results and Discussion

3.1 Run-Time Analysis of the Multi-GPU Framework

For the run-time analysis, five system configurations were investigated, as reported in Table 2. Systems A and B consisted of Tankam et al.: Parallelized multi-graphics processing unit framework for high-speed...



Fig. 2 Multi-graphics processing unit (GPU) architecture for Gabor-domain optical coherence microscopy. The frame grabber is connected to the workstation's motherboard (Supermicro[®]) using PCIe x8. Red color blocks represent the buffering steps. Large arrows represent data transfer of a batch of frames, whereas narrow arrows represent transfer of an individual frame. Double-direction arrows between image processing steps and fused data blocks illustrate the exchange of frames between these two steps. The feedback arrow (Zone # Loop) represents the release of GPUs to handle the next zone. Each of the four GPUs runs independently during the processing and fusing and communicates with the other GPUs during the rendering.



Fig. 3 Illustration of the real-time Gabor fusing process on GPU for one frame.

Table 2	System configurations investigated for Gabor-domain opti-
cal cohei	ence microscopy (GD-OCM) processing.

System	Configuration	Number of graphics processing unit (GPU) Cores
A	2 NVIDIA [®] GTX 680 cards	3072 cores
В	2 NVIDIA [®] GTX Titan	5376 cores
С	4 NVIDIA [®] Tesla C1060	960 cores
D	4 NVIDIA [®] Fermi C2050	1792 cores
E	Intel core i7	

two GTX 680s and two GTX Titans GPUs, respectively, both of which employ adaptive processor and memory overclocking. Systems C and D consisted of four NVIDIA[®] C1060 and C2050 general purpose GPUs, which did not employ adaptive processor and memory overclocking. System E employed an Intel core i7 processor, which provided the benchmark CPU performance.

Table 3 presents the average cumulative run times for the different configurations of Table 2. The run times are presented

Table 3 Computational results using the multi GPU-based GD-OCM image processing for a data volume of 1000×1000 A-scans.

System	Average performance using one GPU (s)	Average performance using two GPUs (s)	Average performance using four GPUs (s)
A	29	13	_
В	31	15	_
С	940	470	235
D	460	230	115

for a single GPU and all GPU scenarios, processing a total of 1000×1000 A-scans. It can be seen that, using System A, a computational time of 29 s with a single GPU and 13 s with two GPUs was obtained. System B provided an average performance of 31 s with a single GPU and 15 s with two GPUs. Compared with a pipelined CPU running time benchmarked at 19 min, Systems A and B produce a computational speedup of $83\times$ and $73\times$, respectively—close to two orders of magnitude speed up. Systems C and D, which employ four GPUs, provided an average computation time of 235 and 115 s, respectively. Their lower performance is attributed to a lower number of processing cores in the system.

Figure 4 presents the average cumulative run time results for the image processing using configurations A and B. For both systems, the performance of the GPUs improved as the number of parallel A-scans being processed was increased. The overall computation time decreased exponentially with an increase in the number of parallel A-scans. For the case in which a total of 8k A-scans were processed in parallel, System A provided an average computation time of 175 and 89 s for the one and two GPU setups, respectively, whereas System B provided computation times of 156 and 82 s.

The results show the scalability of the proposed framework, which is critical for the improvement of the computational speed. From Fig. 4, it can be observed that the computational time taken by two GPUs to perform a given number of A-scans in parallel is equal to around half the computation time taken by a single GPU to perform the same amount of A-scans. Such an observation supports the fact that the framework is agnostic to the number of GPUs used for the computation and thus provides a scalable computational time. In comparing Systems A and B, as the number of parallel A-scans is increased up to 40k, the performance of the two systems is quite comparable. The slight improvement in the performance of the GTX 680 compared with GTX Titan, which has almost twice the number of processing cores, can be attributed to the adaptive processor overclocking achieved by the GPU boost algorithm on the NVIDIA[®] GTX 680 cards.35

As the GD-OCM setup was limited by the camera acquisition time (14.3 s per zone), the number of parallel A-scans



Fig. 4 GPU boost comparison for Systems A and B as a function of the total number of A-scans processed in parallel.

1 Zones (4096 × 1000 × 1000)	Reading data	Data transfer host->device	DC removal	K-space linearization	FFT	Gray level and log scaling	Auto- synchronization	Fusing	Saving	Total time (s)
Pipelined CPU time (s)	Running i the next o	n parallel with operation block	148	846	137	Running in para	allel with previous	operatio	n block	1131
Parallelized two-GTX 680 s time(s)	NA	1.57	2.12	0.57	1.57	3.80	0.67	3.34	NA	13.64

Table 4 Computational speed-up of the multi GPU based GD-OCM image processing compared with a pipelined CPU implementation.

processing was set to 40k for System A to yield a processing time that was faster than the acquisition time. Nevertheless, the implementation presented here leads to a scalable realtime GD-OCM image processing with a finite increase in the number of GPUs used in computation. Table 4 shows the detailed processing time for one zone $(1000 \times 1000 \text{ A-scans})$ on System A with 40k total parallel A-Scan calls per iteration.

The parallelized two-GTX 680s configuration of System A provided a computational time of 13.64 s for one zone $(1000 \times 1000 \text{ A-Scans})$, yielding a computational speed of 600 MB/s. As compared with the pipelined CPU implementation, this gain corresponds to a computational speed-up of over one order of magnitude.

The ability of the proposed framework to account for a larger number of parallel A-scans can be extrapolated from trends observed in Fig. 4; it can be seen that, based on the required computation time, the optimal number of A-scans to be processed in parallel can be selected. As multiple cards are being considered, thermal management of the cards is needed to avoid over-heating, which could lead to a failure of the system-an important consideration for use of the system in a clinical environment where a repeated usage for imaging the patient's anatomy is needed. Instability was experienced on GPU performance when the temperature of the cards went over ~55°C. This occurred when the number of parallel GPU calls was increased. Effective thermal dissipation design that optimally removes heat from all the GPUs, while maintaining the PCs form factor, is critical. As an example, new PC form factors with unified thermal dissipation mechanisms for effective heat removal in GPUs have been investigated by Apple Inc., Cupertino, California⁶⁶ In addition, recent developments in NVIDIA GPUs (Tegra K1) have focused on using a significantly reduced power source while maintaining optimal performance as a means to address GPU over-heating.⁶⁷ Although such designs are in their initial stages of development, future advancements in PC form factors and GPU designs will address the over-heating issue effectively.

The proposed multi-GPU computational framework offers the opportunity to reduce the processing time by the optimal choice of number of parallelization and GPU cards while preserving long-term use of the system. Also, a fully integrated C++ interface is considered to optimize the overall system robustness.

3.2 Processing-Based Image Quality Analysis

The image quality of CPU- and GPU-based processing of the same A-scan was investigated at different steps of the processing to ensure that the same algorithm was implemented on CPU and GPU. For each A-scan the output arrays of each step with CPU and GPU were compared; no difference was found. Figure 5 shows the 2-D image of the same B-scan processed on CPU and on GPU with neither visual nor statistical difference.

The proposed GPU framework was used to image the pointer fingertip. Figure 6 presents three orthogonal views as well as a 3-D volume of the processed and rendered volume using the proposed GPU framework. The cross sectional images of x-z and y-z planes [Figs. 6(a) and 6(c)] show the different layers of the skin (SC: stratum corneum, SG: stratum granulosum, SS: stratum spinosum, and SB: stratum basale). The en face images of x - y plane [Figs. 6(d) and 6(e)] show the granulosum cells nuclei and blood vessels, respectively, demonstrating the high-lateral resolution of the imaging system. Figure 6(b) shows a snap shot of the 3-D volume of 1 mm by 1 mm by 0.6 mm rendered on GPU using max intensity renderer.



Fig. 5 Image comparison between CPU and GPU processing of the same B-scan (zone 3 before the fusing). (a) CPU-based image processing, (b) GPU-based image processing. The depths reported are the distances in skin tissue (n = 1.4). The GPU image was acquired with the architecture A (Two GTX 680).



Fig. 6 GPU-based three-dimensional (3-D) image of the pointer fingertip acquired with the architecture A (Two GTX 680); (a) and (c) represent the cross sectional images of x-z and y-z planes and show the different layers of the skin (SC: stratum corneum, SG: stratum granulosum, SS: stratum spinosum, and SB: stratum basale); (d) and (e) show en face images of x-y plane at two different depths; plane (d) at around the stratum granulosum layer shows the cells nuclei whereas blood vessels can be observed on the plane (e) just below the stratum basal; (b) snap shot of the 3-D volume of 1 mm by 1 mm by 0.6 mm shows the sweat ducts.

4 Conclusion

A scalable and parallelized multi-GPU processing framework was proposed to overcome the processing speed limitation of GD-OCM. Five different scenarios of multi-GPU configurations were tested, and the use of two GTX 680 cards was found to yield the best performance for this application. For one zone $(1000 \times 1000 \times 4096)$, an average performance of ~13 s when processing 40k A-scans in parallel on both cards was achieved, yielding a processing speed of 600 MB/s. This enables realtime processing of a skin volume of $1 \times 1 \times 0.6$ mm³ with 2- μ m resolution. In particular, the goal of reducing the processing time to be faster than the acquisition time was achieved. Over one order of magnitude computational speed-up was obtained compared with the pipelined CPU processing, with no quantitative loss of image information. Importantly, results show that if two GPUs are considered the clinician can visualize the 3-D volume 13 s after the acquisition is completed, and that time is estimated at 6.5 s with four GPUs based on the demonstrated scalability of the framework. Thus, the proposed framework enables real-time processing, a fundamental step on the path toward adoption of GD-OCM in a clinical environment.

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References

- W. Drexler et al., "In vivo ultrahigh-resolution optical coherence tomography," Opt. Lett. 24(17), 1221–1223 (1999).
- V. R. Korde et al., "Using optical coherence tomography to evaluate skin sun damage and precancer," *Lasers Surg. Med.* 39(9), 687–695 (2007).
- J. Welzel et al., "OCT in dermatology," in *Optical Coherence Tomography*, W. Drexler and J. Fujimoto, Eds., pp. 1103–1122, Springer, Berlin, Heidelberg (2008).
- R. Pomerantz et al., "Optical coherence tomography used as a modality to delineate basal cell carcinoma prior to Mohs micrographic surgery," *Case Rep. Dermatol.* 3(3), 212–218 (2011).
- M. Mogensen et al., "OCT imaging of skin cancer and other dermatological diseases," J. Biophotonics 2(6–7), 442–451 (2009).
- F. G. Bechara et al., "Histomorphologic correlation with routine histology and optical coherence tomography," *Skin Res. Technol.* 10(3), 169– 173 (2004).
- M. C. Pierce et al., "Advances in optical coherence tomography imaging for dermatology," J. Invest. Dermatol. 123(3), 458–463 (2004).
- R. Steiner, K. Kunzi-Rapp, and K. Scharffetter-Kochanek, "Optical coherence tomography: clinical applications in dermatology," *Med. Laser Appl.* 18(3), 249–259 (2003).

- A. Alex et al., "Multispectral *in vivo* three-dimensional optical coherence tomography of human skin, *J. Biomed. Opt.* 15(2), 026025 (2010).
- L. E. Smith et al., "Evaluating the use of optical coherence tomography for the detection of epithelial cancers in vitro," *J. Biomed. Opt.* 16(11), 116015 (2011).
- B. J. Vakoc et al., "Cancer imaging by optical coherence tomography preclinical progress and clinical potential," *Nat. Rev. Cancer* 12(5), 363–368 (2012).
- R. Wessels et al., "Optical biopsy of epithelial cancers by optical coherence tomography (OCT)," *Lasers Med. Sci.*, 1–9 (2013).
- J. M. Olmedo et al., "Correlation of thickness of basal cell carcinoma by optical coherence tomography *in vivo* and routine histologic findings: a pilot study," *Dermatol. Surg.* 33(4), 421–426 (2007).
- T. Gambichler et al., "In vivo optical coherence tomography of basal cell carcinoma," J. Dermatol. Sci. 45(3), 167–173 (2007).
- P. Wilder-Smith et al., "Noninvasive imaging of oral premalignancy and malignancy," J. Biomed. Opt. 10(5), 051601 (2005).
- W. B. Armstrong et al., "Optical coherence tomography of laryngeal cancer," *The Laryngoscope* 116(7), 1107–1113 (2006).
- T. Hinz et al., "Preoperative characterization of basal cell carcinoma comparing tumour thickness measurement by optical coherence tomography 20-MHz ultrasound and histopathology," *Acta Derm. Venereol.* 92(2), 132–137 (2012).
- Z. Ding et al., "High-resolution optical coherence tomography over a large depth range with an axicon lens," *Opt. Lett.* 27(4), 243–245 (2002).
- K.-S. Lee and J. P. Rolland, "Bessel beam spectral-domain high-resolution optical coherence tomography with micro-optic axicon providing extended focusing range," *Opt. Lett.* 33(15), 1696–1698 (2008).
- C. Blatter et al., "Extended focus high-speed swept source OCT with self-reconstructive illumination," *Opt. Express* 19(13), 12141–12155 (2011).
- A. Zlotnik et al., "Improved extended depth of focus full field spectral domain Optical Coherence Tomography," *Opt. Commun.* 283(24), 4963–4968 (2010).
- K. Sasaki et al., "Extended depth of focus adaptive optics spectral domain optical coherence tomography," *Biomed. Opt. Express* 3(10), 2353–2370 (2012).
- D. Hillmann et al., "Holoscopy—holographic optical coherence tomography," *Opt. Lett.* 36(13), 2390–2392 (2011).
- A. Ahmad et al., "Real-time *in vivo* computed optical interferometric tomography," *Nat. Photonics* 7, 444–448 (2013).
- A. D. Aguirre et al., "High-resolution optical coherence microscopy for high-speed," *Opt. Lett.* 28(21), 2064–2066 (2003).
- J. P. Rolland et al., "Gabor domain optical coherence microscopy," *Proc. SPIE* 7139, 71390F (2008).
- P. Bouchal, A. Bradu, and A. G. Podoleanu, "Gabor fusion technique in a Talbot bands optical coherence tomography system," *Opt. Express* 20(5), 5368–5383 (2012).
- S. Murali, K. P. Thompson, and J. P. Rolland, "Three-dimensional adaptive microscopy using embedded liquid lens," *Opt. Lett.* 34(2), 145–147 (2009).
- 29. S. Murali et al., "Assessment of a liquid lens enabled *in vivo* optical coherence microscope," *Appl. Opt.* **49**(16), D145–D156 (2010).
- K.-S. Lee et al., "Cellular resolution optical coherence microscopy with high acquisition speed for in-vivo human skin volumetric imaging," *Opt. Lett.* 36(12), 2221–2223 (2011).
- A. W. Greynolds, "Multi-core and GPU accelerated simulation of a radial star target imaged with equivalent t-number circular and Gaussian pupils," *Proc. SPIE* 8841, 88410F (2013).
- 32. J. Bolz et al., "Sparse matrix solvers on the GPU: conjugate gradients and multigrid," *ACM Trans. Graphics* **22**(3), 917–924 (2003).
- 33. C. J. Thompson, H. Sahngyun, and M. Oskin, "Using modern graphics architectures for general-purpose computing: a framework and analysis," in *Proc. 35th Ann. IEEE/ACM Int. Symp. Microarchitect.* (*MICRO-35*), Istanbul, Turkey, IEEE (2002).
- J. Krüger and R. Westermann, "Linear algebra operators for GPU implementation of numerical algorithms," *ACM Trans. Graphics* 22(3), 908–916 (2003).
- R. Fernando, GPU Gems: Programming Techniques, Tips and Tricks for Real-Time Graphics, Addison-Wesley Professional, Indianapolis, Indiana (2004).

- V. M. Bove et al., "Real-time holographic video images with commodity PC hardware," *Proc. SPIE* 5664, 255–262 (2005).
- N. Masuda et al., "Computer generated holography using a graphics processing unit," *Opt. Express* 14(2), 603–608 (2006).
- A. P. Santhanam et al., "A multi-GPU real-time dose simulation software framework for lung radiotherapy," *Int. J. Comput. Assisted Radiol. Surg.* 7(5), 705–719 (2012).
- Y. Min et al., "A GPU-based framework for modeling real-time 3D lung tumor conformal dosimetry with subject-specific lung tumor motion," *Phys. Med. Biol.* 55, 5137–5149 (2010).
- A. Santhanam et al., "Effect of 4D-CT image artifacts on the 3D lung registration accuracy: a parametric study using a GPU-accelerated multi-resolution multi-level optical flow," *J. Med. Phys.* 40(6), SU-E-J-73 (2013).
- Y. Watanabe et al., "Real-time processing for full-range Fourier-domain optical-coherence tomography with zero-filling interpolation using multiple graphic processing units," *Appl. Opt.* 49(25), 4756–4762 (2010).
- K. Zhang and J. U. Kang, "Graphics processing unit accelerated nonuniform fast Fourier transform for ultrahigh-speed, real-time Fourierdomain OCT," *Opt. Express* 18(22), 23472–23487 (2010).
- U.-S. Jung et al., "Simple spectral calibration method and its application using an index array for swept source optical coherence tomography," *J. Opt. Soc. Korea* 15(4), 386–393 (2011).
- K. Zhang and J. U. Kang, "Real-time numerical dispersion compensation using graphics processing unit for Fourier-domain optical coherence tomography," *Electron. Lett.* 47(5), 309–310 (2011).
- 45. Y. Jian, K. Wong, and M. V. Sarunic, "Graphics processing unit accelerated optical coherence tomography processing at megahertz axial scan rate and high resolution video rate volumetric rendering," *J. Biomed. Opt.* **18**(2), 026002 (2013).
- M. Sylwestrzak et al., "Four-dimensional structural and Doppler optical coherence tomography imaging on graphics processing units," *J. Biomed. Opt.* **17**(10), 100502 (2012).
- N. H. Cho et al., "High speed SD-OCT system using GPU accelerated mode for *in vivo* human eye imaging," *J. Opt. Soc. Korea* 17(1), 68–72 (2013).
- Y. Huang and J. U. Kang, "Real-time reference A-line subtraction and saturation artifact removal using graphics processing unit for highframe-rate Fourier-domain optical coherence tomography video imaging," *Opt. Eng.* 51(7), 073203 (2012).
- H. Jeong et al., "Ultra-fast displaying spectral domain optical doppler tomography system using a graphics processing unit," *Sensors* 12(6), 6920–6929 (2012).
- K. K. C. Lee et al., "Real-time speckle variance swept-source optical coherence tomography using a graphics processing unit," *Biomed. Opt. Express* 3(7), 1557–1564 (2012).
- J. Li et al., "Performance and scalability of Fourier domain optical coherence tomography acceleration using graphics processing units," *Appl. Opt.* 50(13), 1832–1838 (2011).
- J. Rasakanthan, K. Sugden, and P. H. Tomlins, "Processing and rendering of Fourier domain optical coherence tomography images at a line rate over 524 kHz using a graphics processing unit," *J. Biomed. Opt.* 16(2), 020505 (2011).
- F. Köttig et al., "An advanced algorithm for dispersion encoded full range frequency domain optical coherence tomography," *Opt. Express* 20(22), 24925–24948 (2012).
- S. Van der Jeught, A. Bradu, and A. G. Podoleanu, "Real-time resampling in Fourier domain optical coherence tomography using a graphics processing unit," *J. Biomed. Opt.* 15(3), 030511 (2010).
- 55. Y. Wang et al., "GPU accelerated real-time multi-functional spectraldomain optical coherence tomography system at 1300 nm," *Opt. Express* 20(14), 14797–14813 (2012).
- Y. Huang, X. Liu, and J. U. Kang, "Real-time 3D and 4D Fourier domain Doppler optical coherence tomography based on dual graphics processing units," *Biomed. Opt. Express* 3(9), 2162–2174 (2012).
- 57. Y. Huang et al., "Motion-compensated hand-held common-path Fourier-domain optical coherence tomography probe for image-guided intervention," *Biomed. Opt. Express* **3**(12), 3105–3118 (2012).
- K. Zhang and J. U. Kang, "Real-time intraoperative 4D full-range FD-OCT based on the dual graphics processing units architecture for microsurgery guidance," *Biomed. Opt. Express* 2(4), 764–770 (2011).

- J. U. Kang et al., "Real-time three-dimensional Fourier-domain optical coherence tomography video image guided microsurgeries," *J. Biomed. Opt.* 17(8), 081403 (2012).
- K.-S. Lee et al., "Dispersion control with a Fourier-domain optical delay line in a fiber-optic imaging interferometer," *Appl. Opt.* 44(19), 4009–4022 (2005).
- K.-S. Lee, K. P. Thompson, and J. P. Rolland, "Broadband astigmatismcorrected Czerny-Turner spectrometer," *Opt. Express* 18(22), 23378– 23384 (2010).
- J. P. Rolland et al., "Gabor-based fusion technique for optical coherence microscopy," *Opt. Express* 18(4), 3632–3642 (2010).
- A. Payne and A. G. Podoleanu, "Direct electronic linearization for camera-based spectral domain optical coherence tomography," *Opt. Lett.* 37(12), 2424–2426 (2012).
- M. Jeon et al., "Full-range k-domain linearization in spectral-domain optical coherence tomography," *Appl. Opt.* 50(8), 1158–1163 (2011).
- Z. Hu and A. M. Rollins, "Fourier domain optical coherence tomography with a linear-in-wavenumber spectrometer," *Opt. Lett.* 32(24), 3525–3527 (2007).
- B. Westove, "Apple Mac Pro Review," http://www.itproportal.com/ reviews/hardware/apple-mac-pro-review/[itproportal.com].
- NVIDIA®, NVIDIA Tegra K1, "A new era in mobile computing," http:// www.nvidia.com/content/PDF/tegra_white_papers/Tegra-K1-whitepaperv1.0.pdf[nvidia.com] (2014).

Patrice Tankam is a postdoctoral research associate at the Institute of Optics with joint appointment at the Center for Visual Science, University of Rochester. He received an engineering and MS degree in instrumentation in 2007, and a PhD in optics in 2010 from the University of Le Mans in France. His research interests include digital holography, interferometry, metrology, optical design, ophthalmology, optical coherence tomography, and image processing.

Anand P. Santhanam is currently an assistant professor in the Department of Radiation Oncology, University of California, Los Angeles. His research focus is on developing algorithms and

techniques that cater to the requirements of medicine. Of particular focus is the development and usage of single GPU and multi-GPU accelerated algorithms for 3-D/4-D image processing, model-based lung registration, anatomy deformation modeling, deformation-based elasticity estimation, tumor dosimetry, and lung deformation-based radiotherapy evaluation.

Kye-Sung Lee is a senior scientist at the Center for Analytical Instrumentation Development in the Korea Basic Science Institute. He earned a PhD in optics at the University of Central Florida in 2008. He conducted research in optical imaging for biological, medical, and material specimen in the Institute of Optics at the University of Rochester from 2009 to 2012. He is interested in developing suitable optical systems to analyze various natures' phenomena in biology, chemistry, physics, space etc.

Jungeun Won is an undergraduate, at the University of Rochester. She is working toward her BS degree with a major in biomedical engineering and a minor in optics. She is a research assistant working on optical coherence tomography for diagnosis of skin cancer. She is interested in developing optical diagnosis techniques.

Cristina Canavesi is the co-founder and president of LighTopTech Corp. and a postdoctoral associate under the NSF I/UCRC Center for Freeform Optics at the University of Rochester. She received the Laurea Specialistica degree in telecommunications engineering from Politecnico di Milano, Milan, Italy, and her PhD in optics from the Institute of Optics at the University of Rochester. She worked in the Institute of Optics Lab at Corecom, Milan, Italy, from 2005 to 2007.

Jannick P. Rolland is the Brian J. Thompson professor of optical engineering at the Institute of Optics at the University of Rochester. She directs the NSF-I/UCRC Center for Freeform Optics (CeFO), the R.E. Hopkins Center for Optical Design and Engineering, and the ODALab (www.odalab-spectrum.org). She holds appointments in the Department of Biomedical Engineering and the Center for Visual Science. She is a fellow of OSA and SPIE.